

Microsatellite diagnosis of species within the *Ceratitis fasciventris*, *C. anonae*, *C. rosa* species complex (Diptera: Tephritidae)

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Current taxonomy

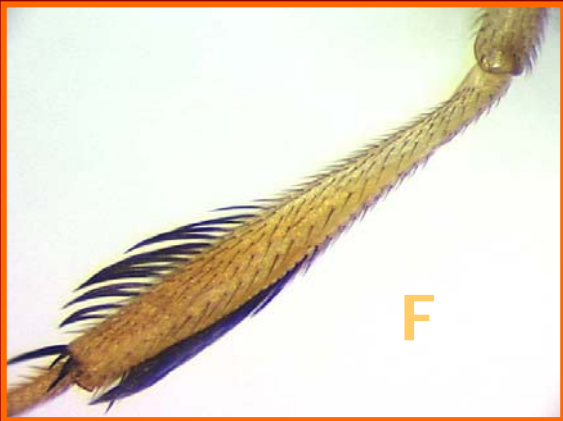
Three recognized species:

- *Ceratitis fasciventris*
- *Ceratitis anonae*
- *Ceratitis rosa*

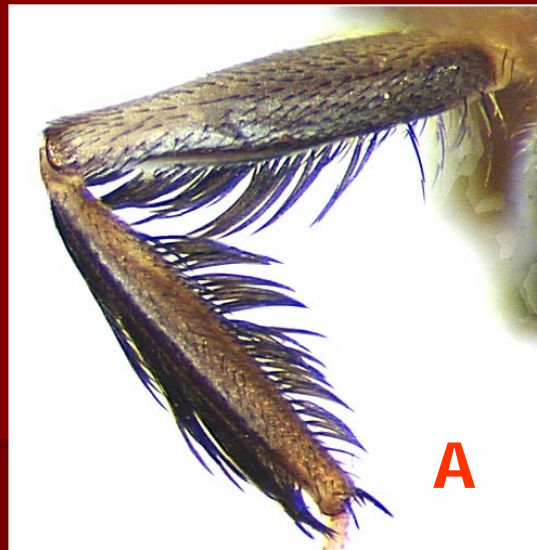


Male differentiation

Based on secondary sexual characters: leg feathering



C. fasciventris



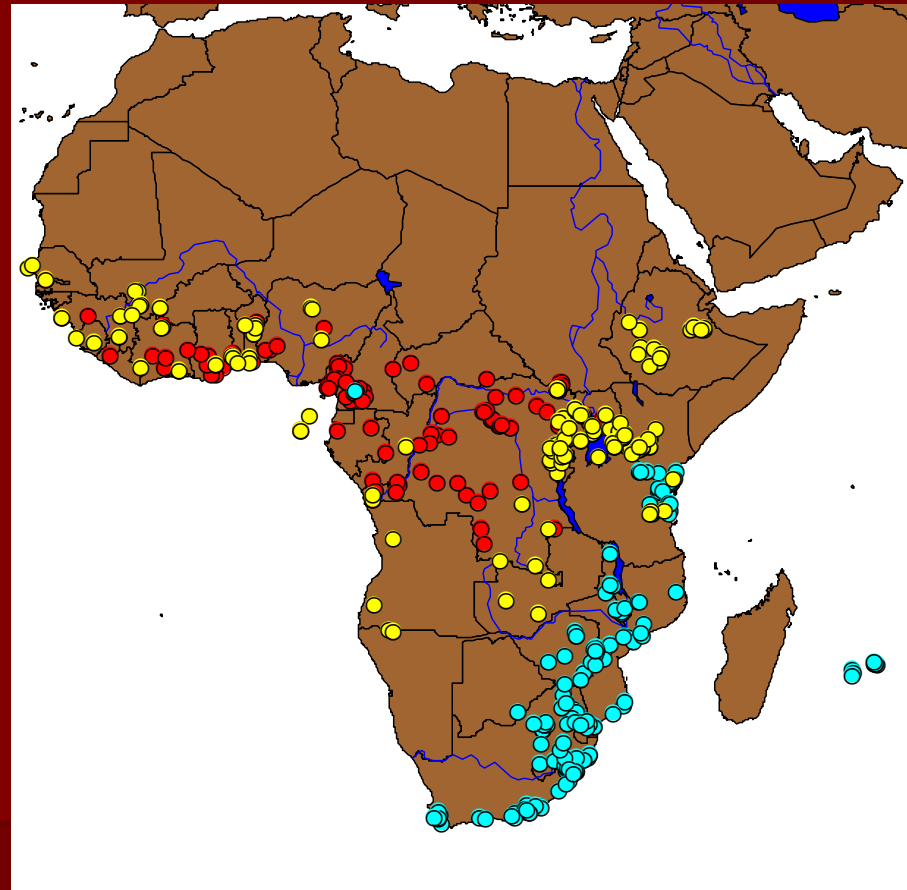
C. rosa

C. anonae

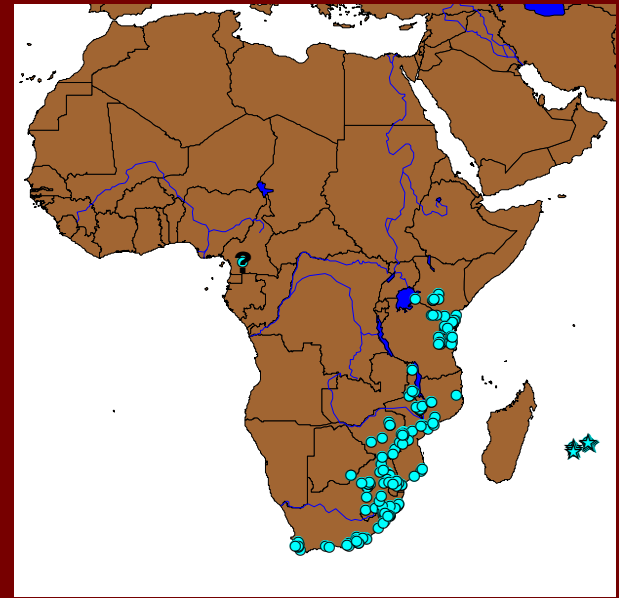
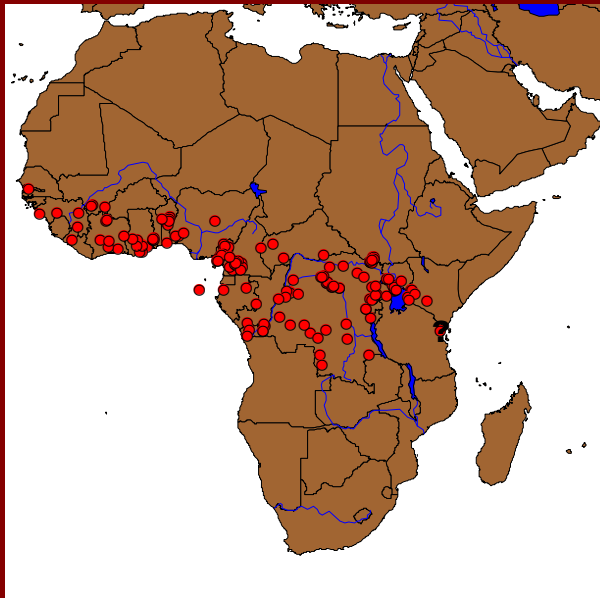
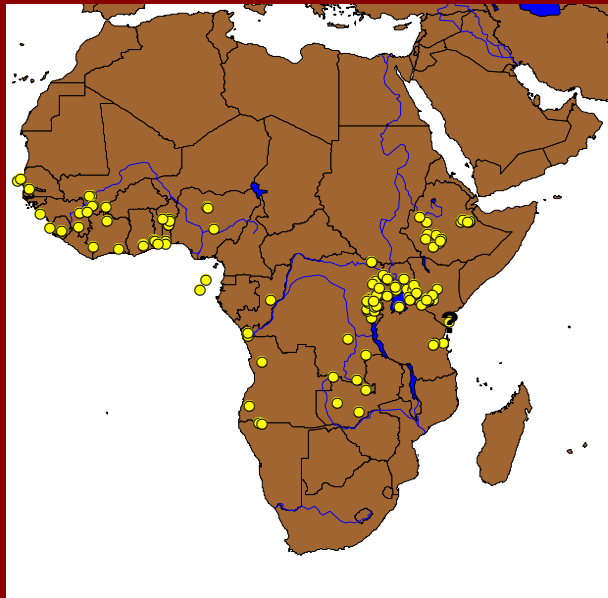


Distribution

Complex widespread
through Africa

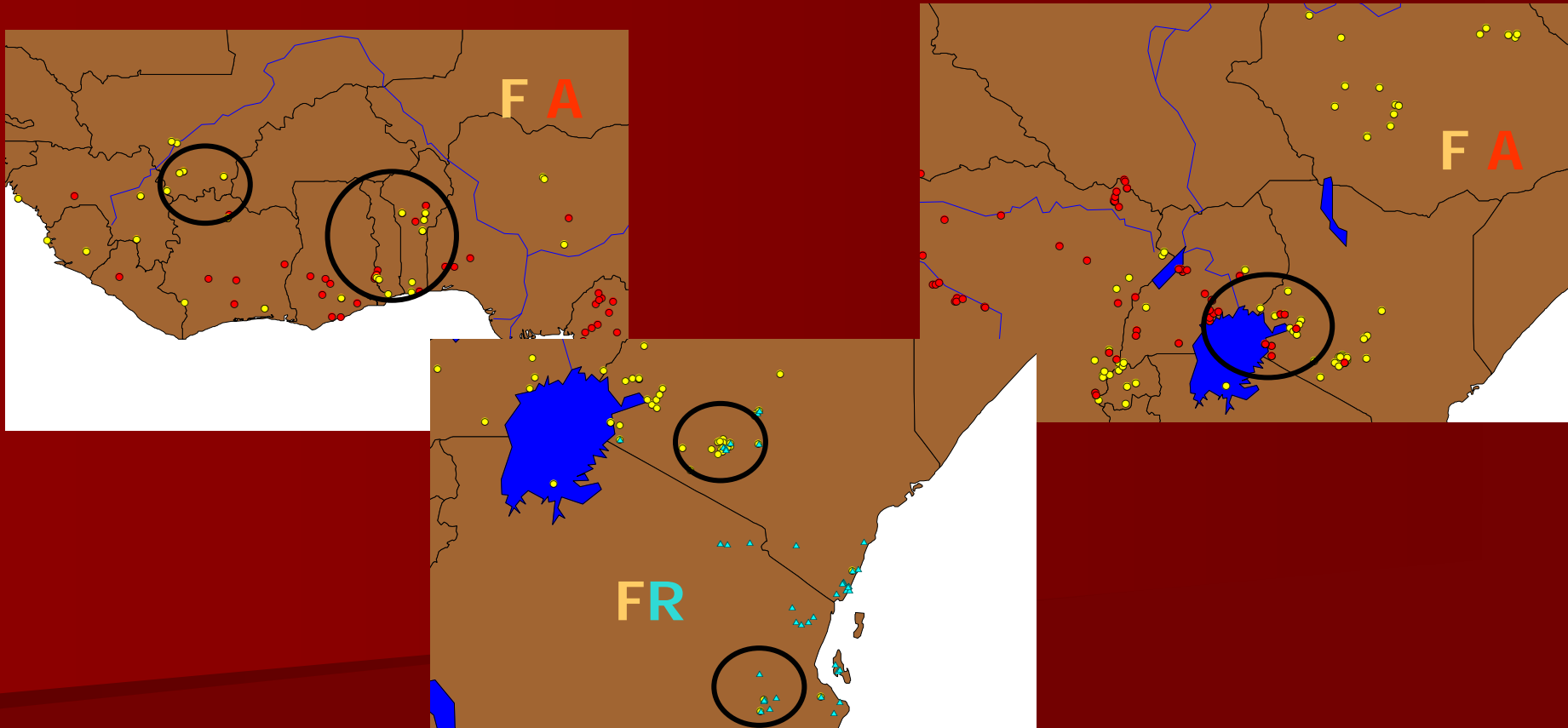


Different distribution patterns



Sympatric or parapatric occurrence

Areas where two species co-occur



Intra-specific variation

Leg coloration male *C. fasciventris*

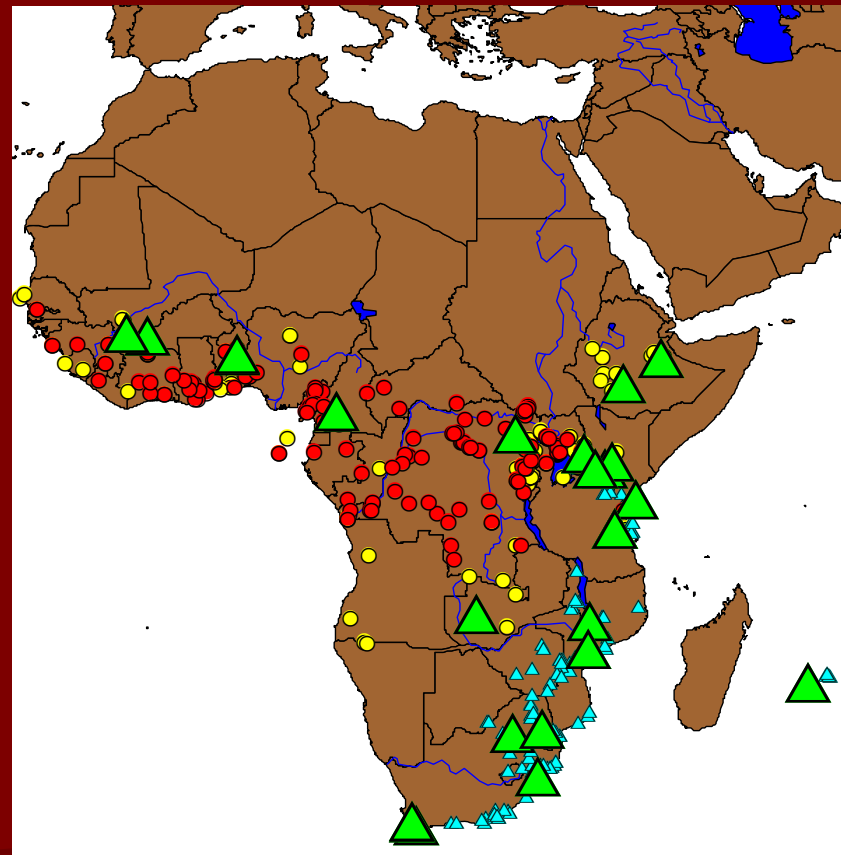
West versus East African populations



Sample selection

27 samples
621 individuals
12 African countries

<i>C. rosa</i>	12 samples, n: 296
<i>C. fasciventris</i>	10 samples, n: 214
<i>C. anonae</i>	5 samples, n: 111



Sample selection

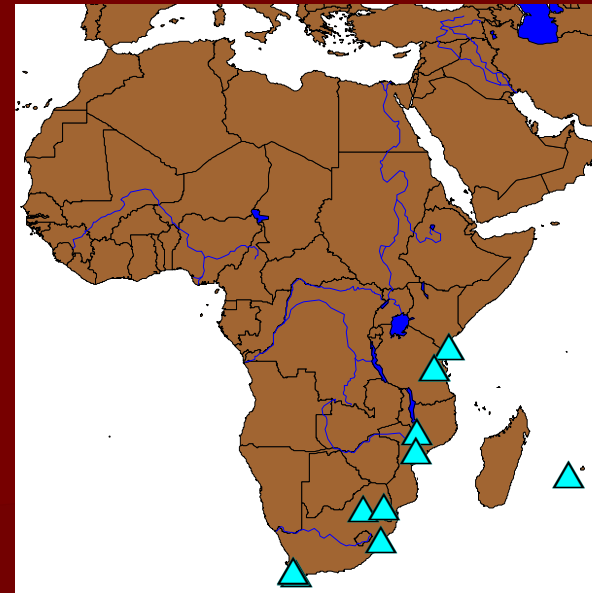
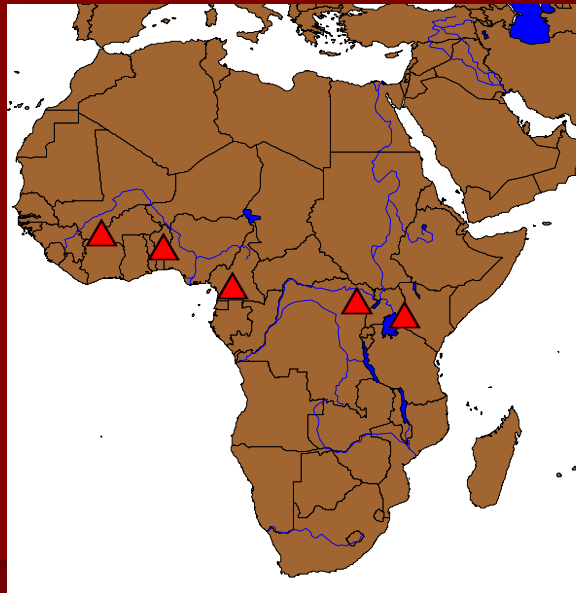
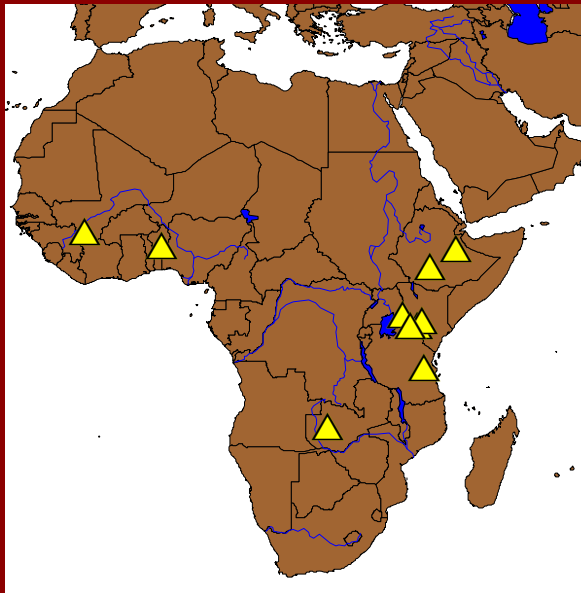
10F/5A/12R

4 sympatric or parapatric
occurrences

sympatric *C. fasciventris* - *C. anonae*
(Kenya - Kakamega forest)

parapatric *C. fasciventris* - *C. anonae*
(Benin – Komin, Benin –Parakon)

sympatric *C. fasciventris* - *C. rosa*
(Tanzania - Morogoro/SUA)



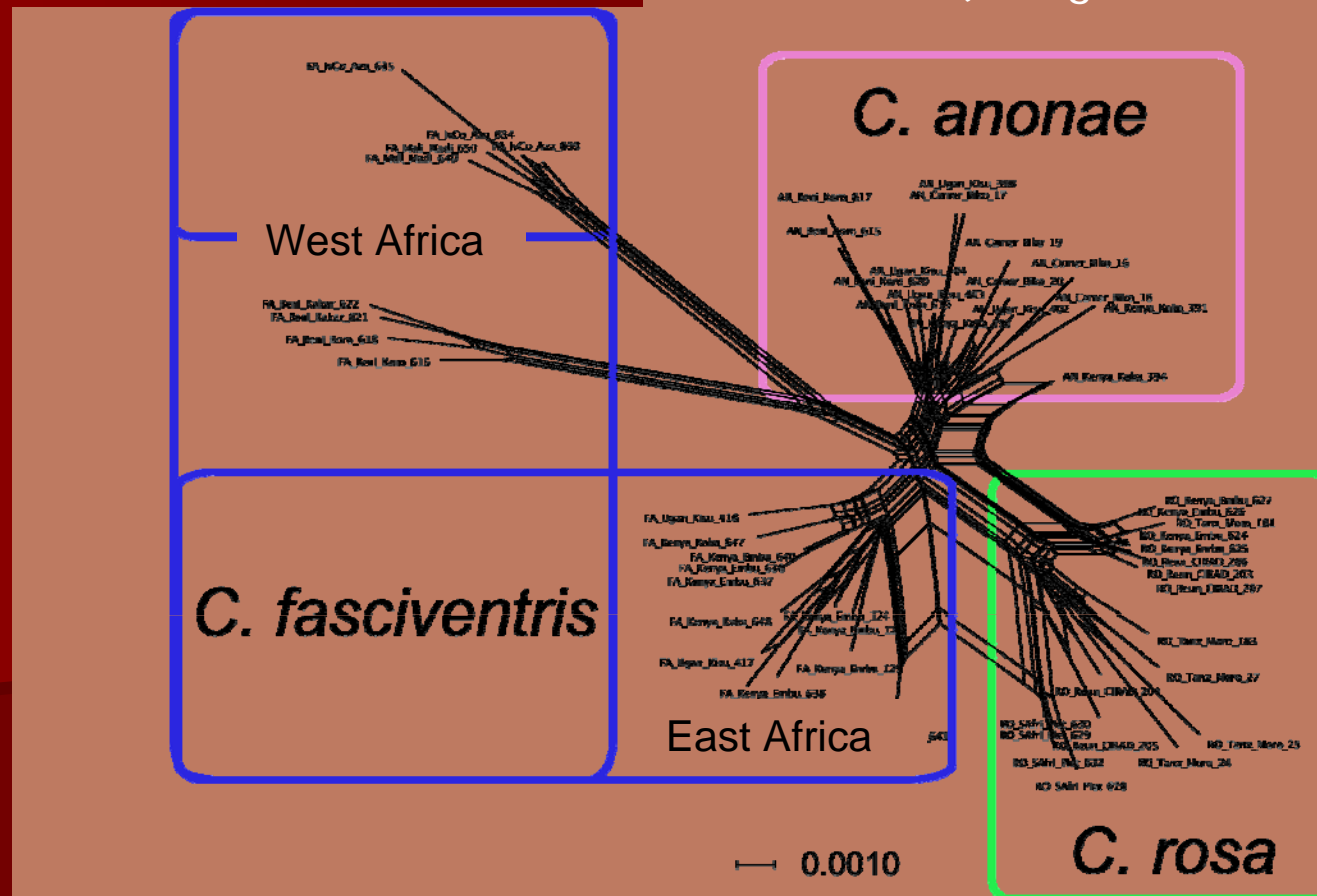
sequence analysis: lack of resolution...

Virgilio, Backeljau, Barr, De Meyer (2008) *Molecular Phylogenetics and Evolution* 48, 270-280

concatenated Per – 16S – COI – ND6

2657 base pairs

NJT network (K2P genetic distance)

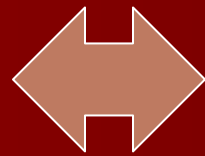
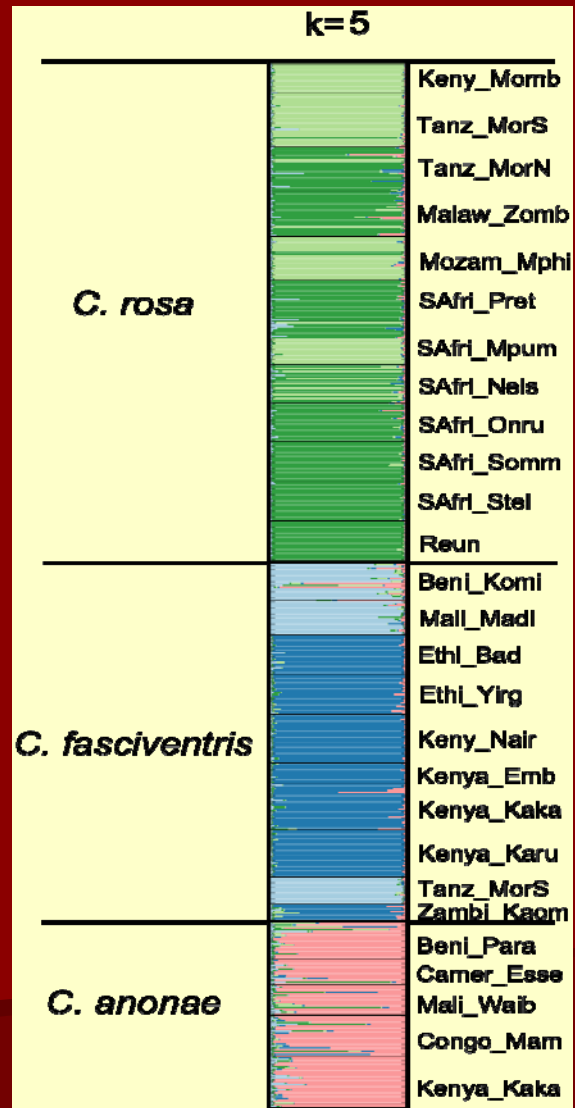


microsatellites

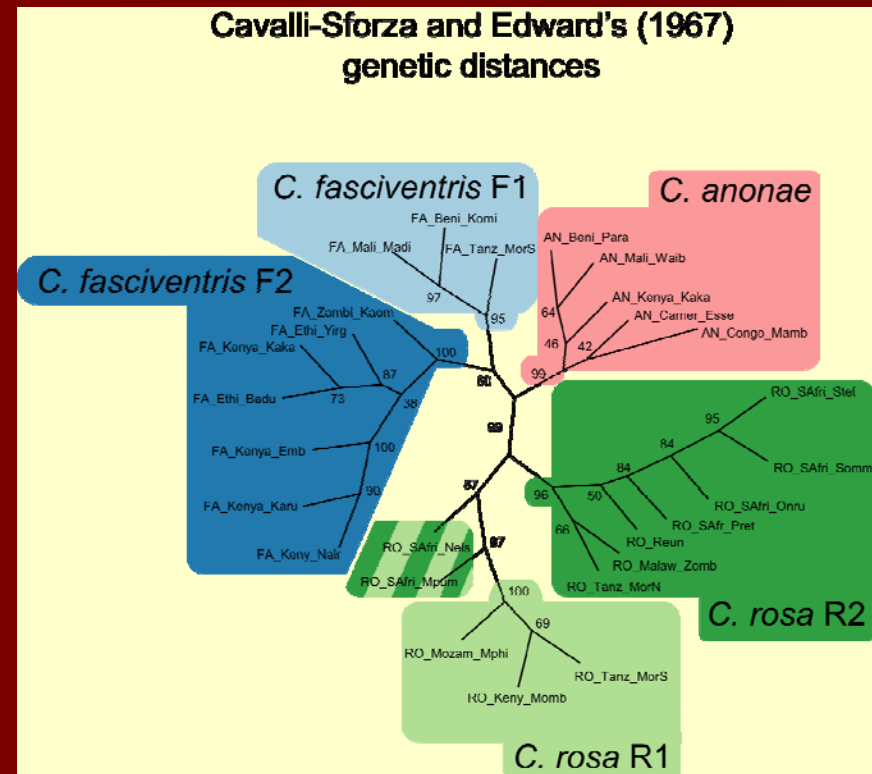
16 loci developed and tested
in collaboration with CIRAD – La Réunion

locus	F pimer	R pimer	dye	repeat	length range
F_1	AGGTTGTGGTATCTACGAAAGG	GCCTGCAAATCTTTAGCAACA	6FAM	caa	70-110
F_2	GAGAAGGTAGGCAACGGATG	TTGCGTTTTGTATACGCTCG	6FAM	atc	160-200
F_3	AGGCATCACATGCACTCCTT	TGGGGAGGCAAGACAAATAA	VIC	ca	80-120
F_4	ATGAAAAAGACACCGCTTGC	AGCGGCATAACCGTAACAAT	VIC	tg	160-200
F_5	TCACAAGACGATATTTAAGTTGCT	GCACCTGCACTTGTCACT	PET	tgt	70-100
F_6	ATATTCCCCAATTCAGTGCC	TTGCACAAGTACCAGTTCTACGA	PET	tg	250-300
F_7	AAATGCAGATAATGCAAGCG	TCAACAGCTTTGCTCTCAGTG	NED	ga	80-110
F_8	AAATCAAACACGGTTACAACCAC	ATCTAATTCTTCTTTTCGACGGC	NED	caa	230-300
F_9	TTTGAGTCTCCGAAAGCAGC	CGAAATAGAGCCAAATCGAA	6FAM	ttg	100-150
F_10	CCAGCCACCAACAGTTTTCT	TTTGTGTGGGTGGCACTTTA	6FAM	aca	195-250
F_11	TTCCTCTTACACCACCAGCC	CGTCTGTCGTCATTCCGTTA	VIC	atgt	90-120
F_12	TTATTTTACTTGCGTCCGCTT	GTATGTCCAGTACGAGGGCG	VIC	tgta	170-200
F_13	TTTACCGTTGCCACTTTTCA	AACTGCAACCAGCAGAGTCA	PET	atc	140-210
F_14	TATCTGCCTTCCGAAGCTC	CGTCGTCGATTCTACGGACTA	PET	ac	260-290
F_15	CTTGTGGCAAGCAAGTGAAG	AAATGTGCTATAAATGGCTATGG	NED	gtat	80-130
F_16	CAAATAAATTCCTTCTTCTTCTTC	GTGTTGTTGCCGTTGCTTC	NED	cac	220-300

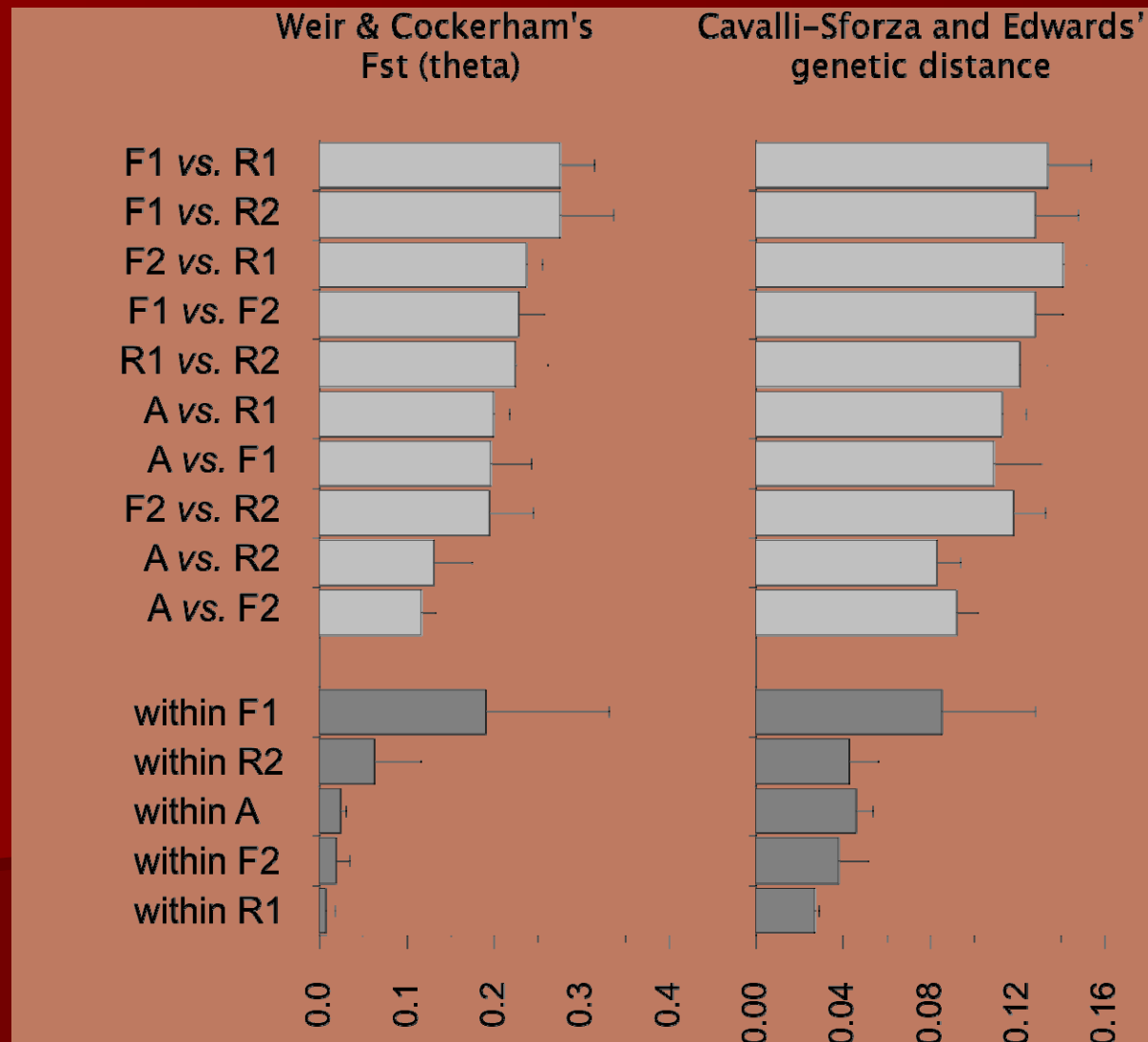
individual assignments



population distances

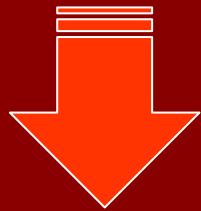


average genetic differentiation
among types R1, R2, F1, F2, A



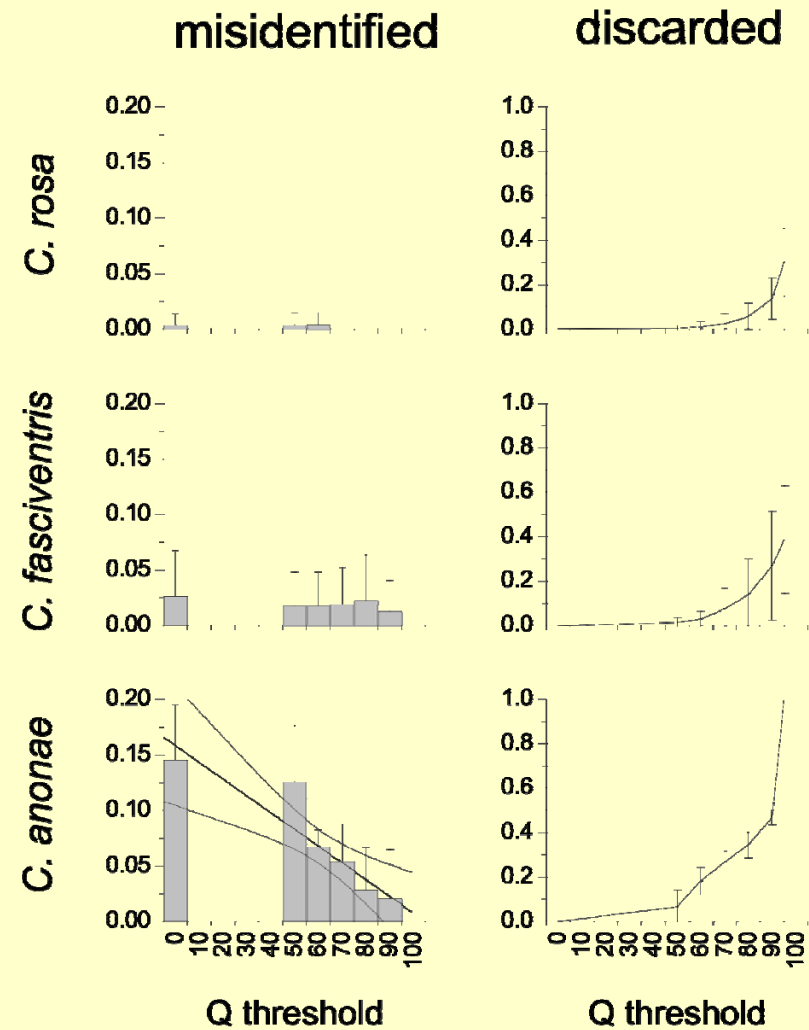
microsatellites diagnosis
of FAR morphospecies:

R1 or R2 -> *C. rosa*
F1 or F2 -> *C. fasciventris*
A -> *C. anonae*



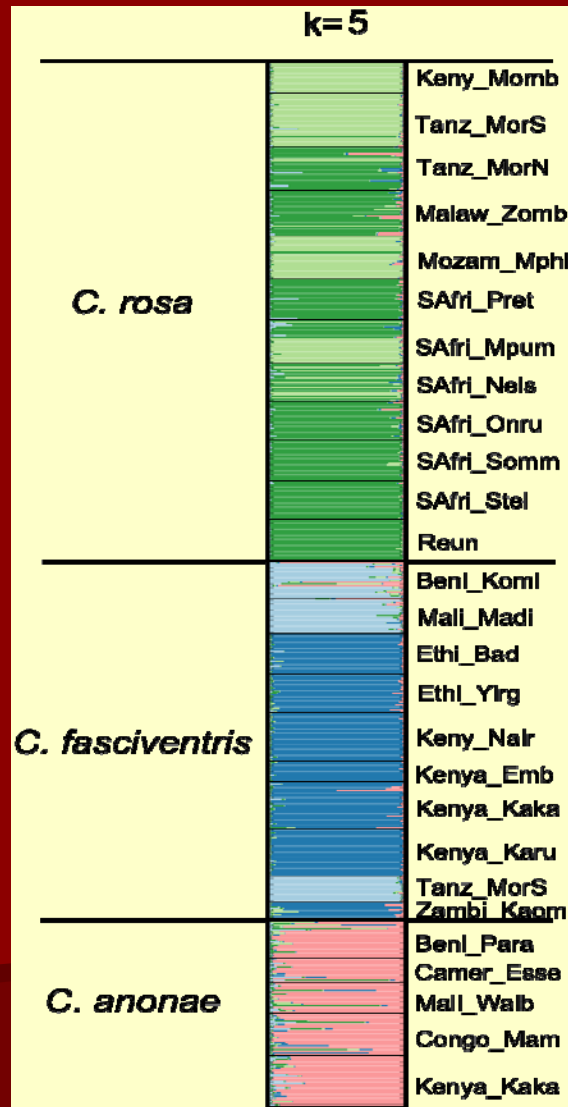
do we need an ID threshold?

molecular diagnosis FAR



proportions of misidentified and discarded specimens
averaged across populations (SD as error bars)

genetic- and morpho-types



genetic types

R1

R2

F1

F2

A

morphotypes

males

females

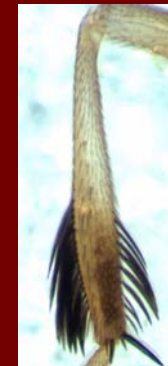
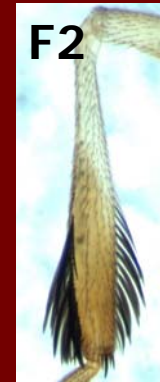
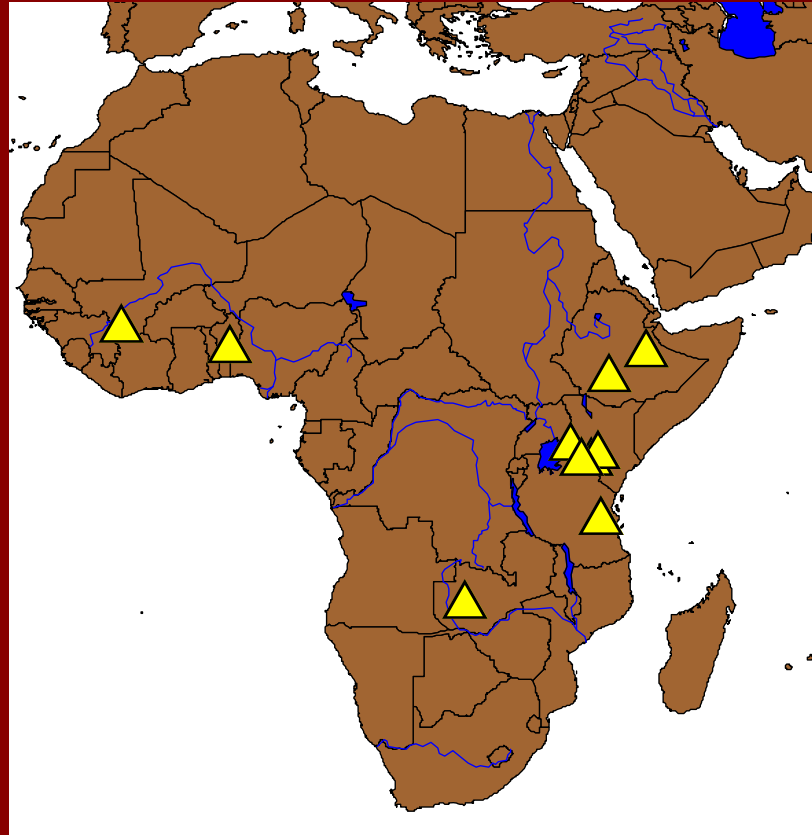


?

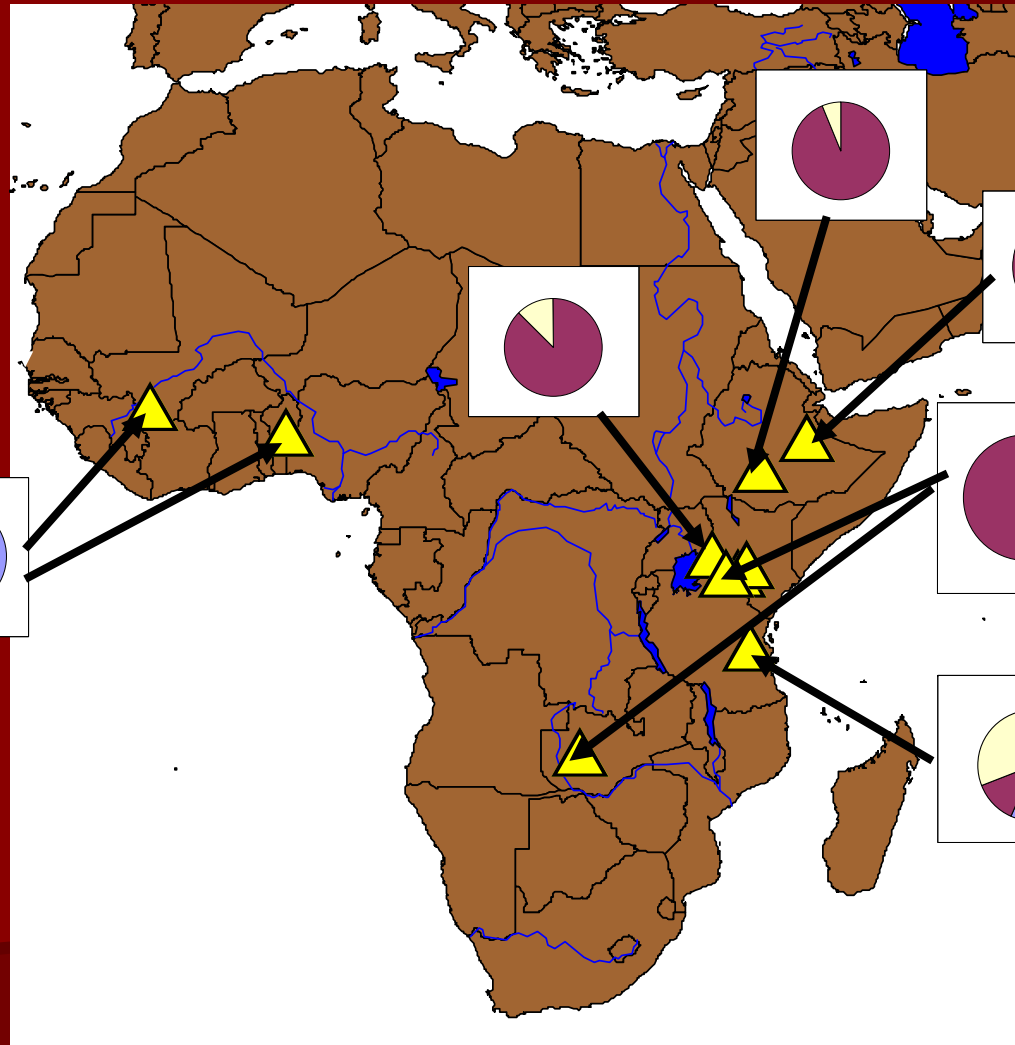
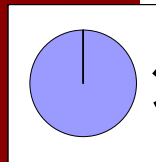
?



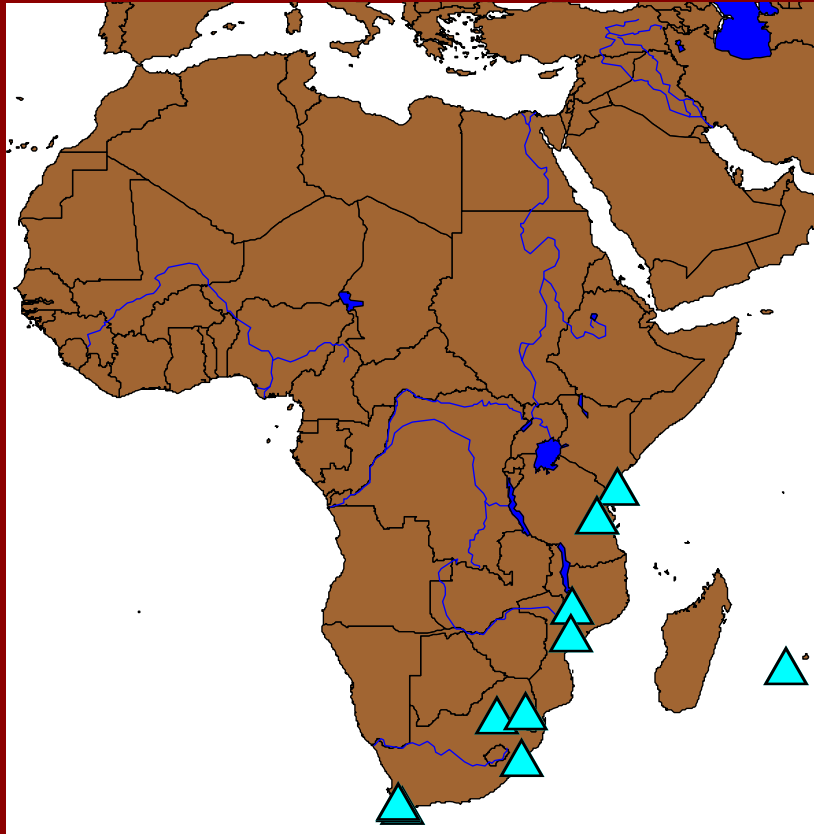
Morphotypes *C. fasciventris*



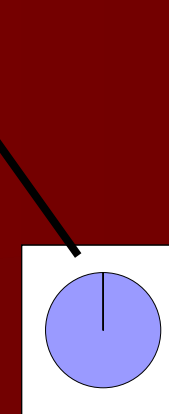
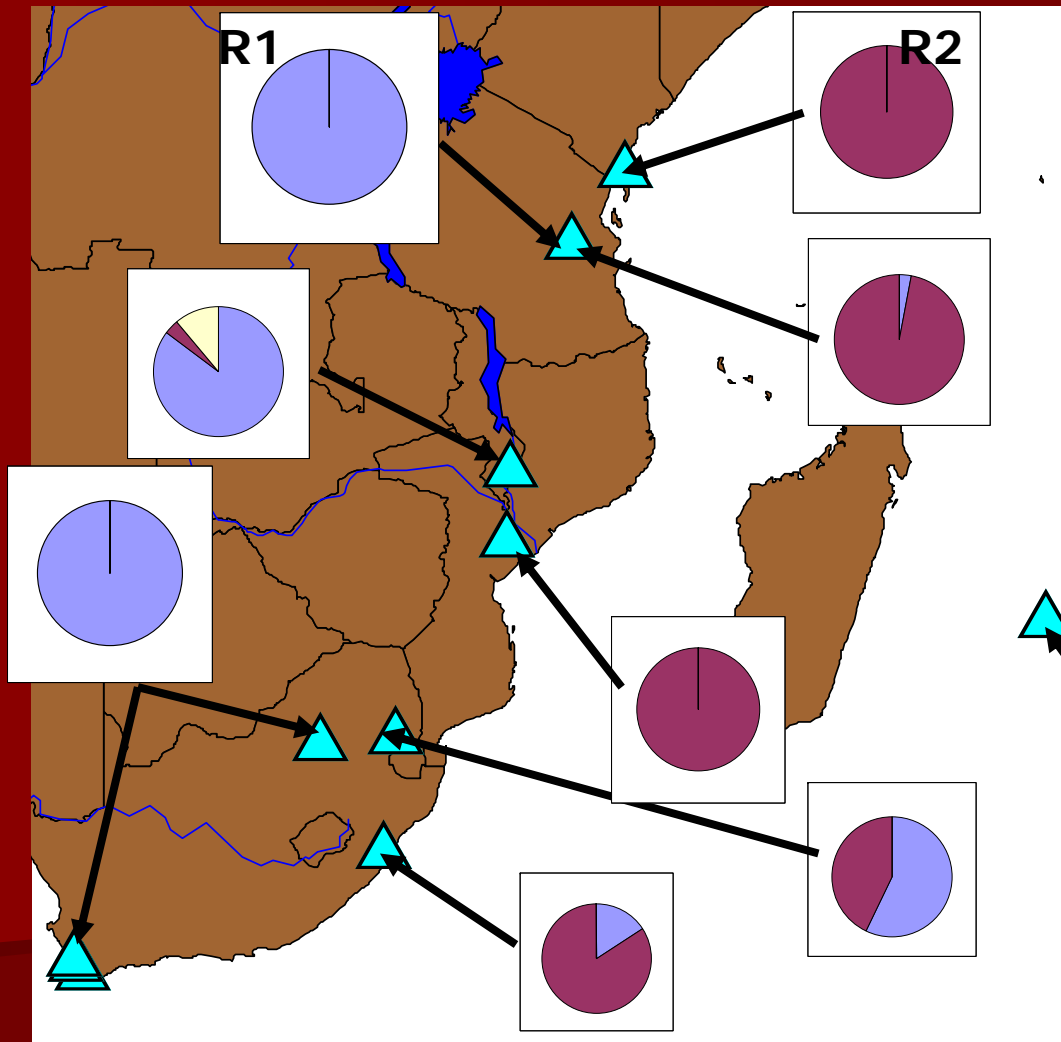
Morphotypes *C. fasciventris*



Morphotypes *C. rosa*



Morphotypes *C. rosa*



Biotypes *C. rosa* ?

Studies on developmental rates

Duyck & Quilici (2002): *C. rosa* has lower minimum temperature thresholds than *C. capitata*. Based on culture originating from Réunion Island

Duyck et al (2006): climatic niche partitioning with *C. rosa* preferring humid and intermediate to cold areas (whereas *C. capitata* prefers dry and intermediate to warm areas)

Grout & Stoltz (2007): *C. rosa* results differ substantially from those of Duyck et al., with higher minimum temperature threshold for development. Based on culture originating from Nelspruit supplemented with specimens from Mpumalanga Province.



Biotypes *C. rosa* ?

Réunion population corresponds to R1 type, while Nelspruit and Mpumalanga populations are partially or largely R2 type.

Tanzania: Morogoro population (400-500 masl) correspond largely to R2 type while Nyandira population (1600 masl) correspond to R1 type.

ICIPE colony corresponds to R1 type. Originates from Central Highlands. Mombasa population corresponds to R2 type.



Biotypes *C. rosa* ?

Need for further study: do the two types of *C. rosa* that are differentiated genetically and morphologically, correspond with two biotypes that have different climatic requirements regarding development?

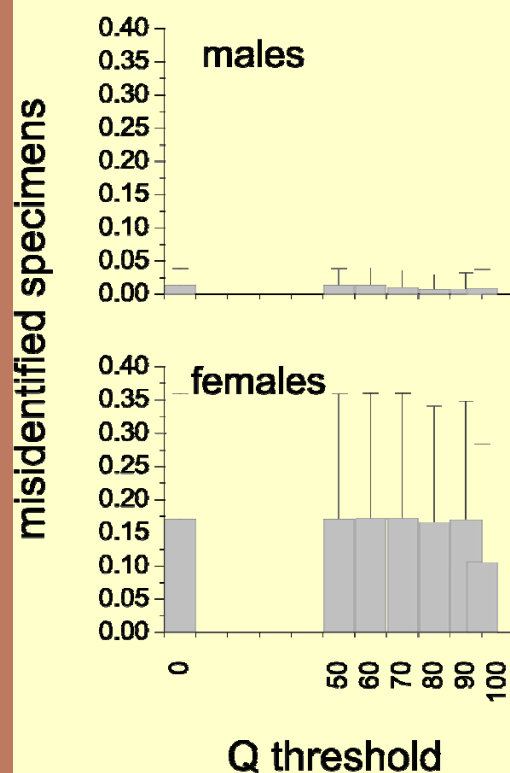
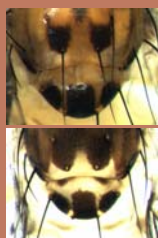
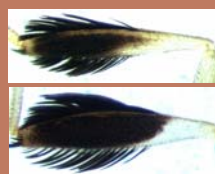
Experiment under controlled conditions should be repeated with identical conditions (same climatic chamber) and specimens studied genetically and morphologically.

If confirmed, implications for control and invasive power

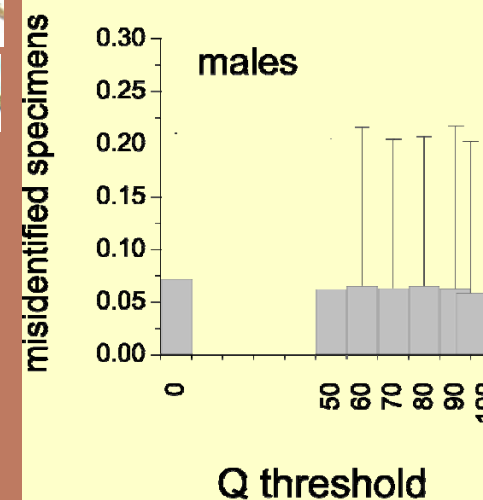


morphological diagnosis of genetic types R1, R2 (males and females) and F1, F2 (males)

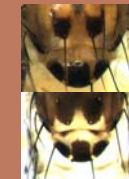
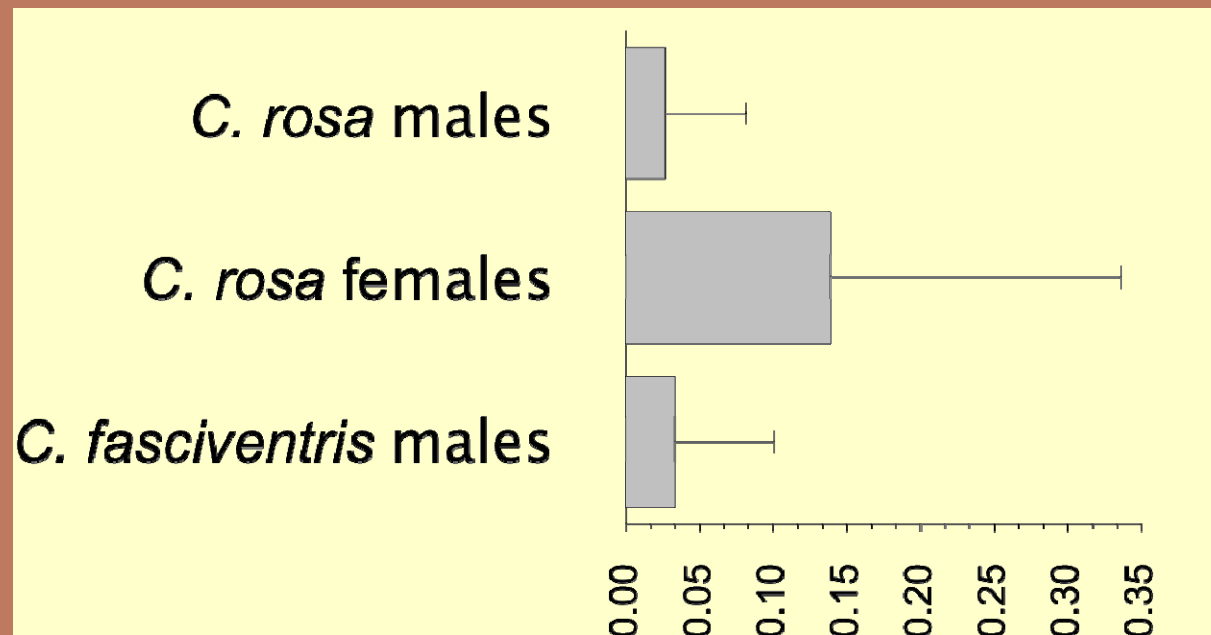
morphological diagnosis of R1, R2 (*C. rosa*)



morphological diagnosis of F1, F2 (*C. fasciventris*)



uncertain morphological diagnosis
of types R1, R2, F1, F2



Summary

Micorsatelite diagnosis of FAR morphospecies (males and females)

- *C. rosa*: low ID error (<5%), highly consistent results across populations
- *C. fasciventris*: low ID error (<5%), consistent results across populations
- *C. anonae*: ID error dependent on Q threshold



Summary

Morphological diagnosis of R1, R2, F1, F2 types

- *C. rosa* males: low ID error (<5%), consistent results across populations
- *C. rosa* females: higher ID error (<10%), not consistent results across populations
- *C. fasciventris*: males ID error >15%, not consistent results across populations

proportion of morphologically-not-identified types:

C. rosa males < *C. fasciventris* males < *C. rosa* females



Future research

- Morphological characters to separate females within the FAR complex: wing landmarking
- *C. rosa* biotypes: climate chambers experiments
- Genotypes *C. fasciventris* also different biotypes?
- Differences host spectrum?
- Ecological niche modeling: data need re-analysis
- Links larval morphology, cuticular hydrocarbons



Acknowledgements

- IAEA for organizing the CRP and financial support participation of MDM
- Dick Drew & colleagues (Griffith University) for organizing this meeting
- Various collectors and collaborators in Africa for providing samples
- Action 1 Belgian Science Policy: funding research MV
- Joint Experimental Molecular Unit (Tervuren & Brussels)

